



بِسْمِ اللّٰهِ الرَّحْمٰنِ الرَّحِیْمِ

Shendi University

Faculty of Postgraduate Study and Scientific Research



Frequency of Survivin Expression in Breast Cancer using Immunohistochemistry

Thesis submitted for partial fulfillment for the requirement of master degree in medical
laboratory science (Histopathology and Cytology)

By:

Manal Omer Ali Osman

B.Sc. In Histopathology, Cytology and Haematology

Shendi University 2001

Supervisor:

Dr. Ahmed Mohamed Ahmed

2018

الآية

بِسْمِ اللَّهِ الرَّحْمَنِ الرَّحِيمِ

قال تعالى:

**(فَقُلْتُ اسْتَغْفِرُوا رَبَّكُمْ إِنَّهُ كَانَ غَفَّارًا (10) يُرْسِلُ السَّمَاءَ عَلَيْكُمْ
مِدْرَارًا (11) وَيَمْدِّكُمْ بِأَمْوَالٍ وَبَنِينَ وَيَجْعَلُ لَكُمْ جَنَّاتٍ وَيَجْعَلُ لَكُمْ
أَنْهَارًا (12))**

صدق الله العظيم

سورة نوح من الآية (10 - 12)

Dedication

I dedicate this research to my father, my mother, my husband, my sister, my children and my brothers.

Acknowledgment

First and foremost, I thank Allah for letting me live to see this dissertation through. I am for ever indebted to Allah who support and give me power to do this dissertation.

I would like to thank my supervisor Dr. Ahmed Mohamed Ahmed, for his patience, continuous guidance throughout my dissertation with his knowledge.

I am grateful to all my teachers and colleagues in the department of histopathology and cytology, college of medical laboratory science, Shendi University, for their help and support.

Thanks to all my friends in the master program.

Finally, I am grateful to my family for their constant support and encouragement.

Abstract

This is retrospective a laboratory based descriptive study which was conducted at ELrhama medical center during the period from January 2018 to April 2018. The study was aimed to detect survivin antigen expression in breast carcinoma using immunohistochemistry.

Forty paraffin embedded blocks were collected from patients samples previously diagnosed as invasive ductal carcinoma. Grade I not reported, grade II 11(27.5%), grade III 29 (72.5%).

Tissue microarrays prepared from paraffin blocks, were cut by rotary microtome at 3 μ m, and then stained by immunohistochemical method (new indirect technique) .The data obtained was analyzed using SPSS program version 22.

The age of patients ranged between 30 to 71 years with mean age of 48.6 years. Most of patients were less than 50 years representing 23(57.5%) and the remaining 17 (42.5%) were more than 50 years.

Survivin expression was positive in all samples of malignant tumor. Regarding distribution of breast carcinoma samples in relation to tumor grading ,grade II the expression was moderate 11 (27.5%) , grade III was high 29(72.5%) ,this result showed significant association between survivin antigen expression and breast carcinoma (p-value = 0.000).

Regarding the relation between age and expression of survivin antigen was found , less than 50 years 7(17.5%) expression was moderate, 16(40%) high expression, more than 50 years 4(10%) expression was moderate, 13(32.5%) high expression .This result showed no association between survivin antigen expression and age (p-value = 0.508).

The study concluded that the expression of survivin antigen was associated with breast carcinoma. Distribution of survivin expression does not affected by age.

المستخلص

أجريت هذه الدراسة الوصفية التحليلية فى مركز الرحمة الطبى شعبة الأنسجة والخلايا المريضة بولاية الخرطوم فى الفترة من يناير إلى ابريل 2018، هدفت الدراسة للكشف عن ظهور مستضد السيريفيين فى أورام الثدي الخبيثة باستخدام كيمياء الأنسجة المناعية .

جُمع أربعون قالب شمعى من عينات مرضى كانوا مشخصين مسبقاً على أنهم مصابون بأورام الثدي الخبيثة. المرحلة الأولى من المرض لم تسجّل ، المرحلة الثانية من المرض كانت تمثل 11 بنسبة (27.5 %) ، والمرحلة الثالثة من المرض كانت تمثل 29 بنسبة (72.5%) .

أختيرت مصفوفات الانسجة الصغيرة من قوالب الشمع ، وقطعت باستخدام المشراح الدوار بسمك 3 ميكرون ، صبغت بواسطة طريقة كيمياء الانسجة المناعية (الطريقة الجديدة غير المباشرة) ، واستخدم برنامج الحزم الإحصائية للعلوم الاجتماعية ، النسخة 22 لتحليل البيانات.

تراوحت أعمار المرضى بين 30 الى 71 عام بمتوسط عمر 48.6 سنة. أظهرت الدراسة أن معظم المرضى كانت أعمارهم أقل من 50 سنة كان عددهم 23 بنسبة (57.5%) والذين كانت أعمارهم أكثر من 50 سنة 17 بنسبة (42.5%).

أظهرت الدراسة أن مستضد السيريفيين كان موجب الظهور فى جميع العينات مع وجود علاقة ذات دلالة إحصائية بين ظهور المستضد ومراحل الورم (القيمة الاحتمالية = 0.000) .

فيما يتعلق بالعلاقة بين ظهور مستضد السيريفيين وأعمار المرضى وجد أن الأعمار أقل من 50 سنة كانوا 7 وبنسبة (17.5%) ظهر مستضد السيريفيين بشكل متوسط ، 16 و بنسبة (40%) ظهر بشكل قوي ، أما الأعمار اكثر من 50 سنة ظهر مستضد السيريفيين بشكل متوسط في 4 وبنسبة (10%) ، وظهر بشكل قوي في 13 وبنسبة (32.5%) .مع عدم وجود علاقة ذات دلالة إحصائية بين ظهور مستضد السيريفيين وأعمار المرضى (القيمة الاحتمالية = 0.508) .

خلصت الدراسة الى أن ظهور مستضد السيريفيين له ارتباط مع أورام الثدي الخبيثة مع عدم وجود علاقة مع توزيع أعمار المرضى.

List of contents

Contents	Page
الآية	I
Dedication	II
Acknowledgement	III
Abstract(English)	IV
المخلص	VI
List of contents	VII
List of tables	IX
List of abbreviations	X
Chapter one – Introduction	
1.1 Introduction	1
1.2 Objectives	3
Chapter two –Literature Review	
1.2 Breast Anatomy and histology	4
2.2 Breast pathology	5
2.2.3 Tumors of the breast	5
2.3 Epidemiology of breast cancer	10
2.4 Risk factors of breast cancer	11
2.5 Method of diagnosis of breast cancer	12
2.6 Breast cancer treatment	14
2.7 Survivin	13
2.8 Survivin and breast cancer	14
Chapter three-Material and Methodology	
3.1 Material	15

3.2 Methodology	15
3.2.1 Study design	15
3.2.3 Study area	15
3.2.4 Immunohistochemistry staining	15
3.2.5 Data analysis	16
3.2.6 Result interpretation	16
3.2.7 Ethical consideration	16
Chapter four - Result	
4.1 Result	17
Chapter five - Discussion	
5.1 Discussion	22
Chapter six - Conclusion and Recommendations	
6.1 Conclusion	23
6.2 Recommendations	24
References	25
Appendices	31

List of tables

Table No	Title	Page
Table (4.1)	Distribution of breast carcinoma according to patients age	18
Table (4.2)	The grading of breast carcinoma among the study samples	19
Table (4.3)	Correlation between grading and expression of survivin in breast carcinoma	20
Table (4.4)	Correlation between expression of survivin and patients age	21

List of abbreviations

IAP	Inhibitor of Apoptosis
FNA	Fine Needle Aspiration
DCIS	Ductal Carcinoma In-Situ
ELISA	Enzyme-Linked Immunosorbent Assay
PARP	Poly ADP Ribose Polymerase
MRI	Magnetic Resonance Imaging
DAB	Di Amino Benzedrine Tetrahydrochloride
SPSS	Statistical Package for Social Science
DPX	Distyrene a plasticizer and xylene

Chapter One

Introduction

Objectives

1. Introduction

1.1 Introduction:

Breast cancer is the most common cancer in women, is the cancer that arises from breast cells, and contains many types ⁽¹⁾. Breast cancer is the most common cancer in women worldwide, with nearly 1.7 million new cases diagnosed in (2012). This represents about 12% of all new cancer cases and 25% of all cancers in women. It is the fifth most common cause of death from cancer in women. The four most common cancers occurring worldwide are lung, breast, bowel and prostate cancer these four account for around 4 in 10 of all cancers diagnosed Worldwide there will be 23.6 million new cases of cancer each year by 2030 (estimated). Breast cancer occurs when abnormal cells in the breast multiply uncontrollably to form a tumor. While most cases of breast cancer occur in women it does occur in men too, although this is rare (about 1% of cases). Survival rates for breast cancer vary worldwide. Advanced and metastatic breast cancer is currently incurable, but treatable, with a median survival rate of 2-3 years ⁽²⁾.

In Sudan data revealed that 74% of the women were < 50 years old or premenopausal ⁽³⁾.

Changes or mutations in DNA can cause normal breast cells to become cancer. Certain DNA changes are passed on from parents (inherited) and can greatly increase your risk for breast cancer. Other lifestyle-related risk factors, Hormones seem to play a role in many cases of breast cancer, but just how this happens is not fully understood ⁽⁴⁾.

Methods of diagnosis of breast cancer are via fine needle aspirate (FNA), mammography, immunohistochemistry, tissue biopsy, and molecular diagnosis ⁽⁵⁾.

Treatment options include, Surgery, Radiation therapy, Hormone therapy, Chemotherapy, Targeted therapy and may be more than one type of treatment ⁽⁶⁾.

Survivin is a protein that inhibits apoptosis and regulates cell division ⁽⁷⁾. The survivin gene spans 15 kb, and is located on chromosome 17 at band q25 ⁽⁸⁾.

Survivin is a member of the inhibitor of apoptosis (IAP) family. The survivin protein functions to inhibit caspase activation, there by leading to negative regulation of apoptosis or Programmed cell death ⁽⁹⁾. Several studies were carried out to assess the possibility of survivin as a prognostic molecule. For example, in a study of 275 patients with breast cancer (70), survivin mRNA was highly expressed in tissues from younger patients (<50) and in high-grade cancer tissues. High survivin concentrations were most strongly associated with ER- or PR-negative tumors. Survivin demonstrates a strong, independent association with poor prognosis. Survivin might be used as a new marker to stratify breast cancer patients for more optimal treatment modalities, or it could be a promising new target for therapy. In another study, survivin expression was examined in 167 cases of breast cancer and the results suggest that apoptosis inhibition by survivin, is a significant prognostic parameter of worse outcome in breast carcinoma ⁽⁹⁾.

Rationale:-

Survivin is apoptosis inhibitor, and usually expressed in tissue as indicator for cancer cells, so it's expression in breast cancer may be helpful in diagnosis and prognosis for breast cancer detection and monitoring.

1.2 Objectives:

1.2.1 General objective:

To study the expression of survivin in breast cancer among Sudanese women patients.

1.2.2 Specific objectives:

- To detect the expression of survivin in the samples of breast cancer patients.
- To determine the percentage of survivin expression in breast cancer tissue.
- To detect the correlation between the survivin as a tumor marker and the histological grading of breast cancer.

Chapter Two

Literature review

2. Literature review:

2.1 Breast Anatomy and histology:

The breast is a modified skin gland enveloped in fibrous fascia. The superficial pectoral fascia is located just beneath the skin and in the retro mammary space. The undersurface of the breast lies on the deep pectoral fascia. Although there are fascial layers between the breast proper and the pectoralis major muscle, the breast is not completely separate from the pectoralis major muscle, as there are penetrating lymphatics and blood vessels. The breast is composed of three major structures: skin, subcutaneous tissue, and breast tissue (parenchyma and stroma). The parenchyma is divided into 15 to 20 lobes or segments that converge at the nipple in a radial arrangement. The ducts from the lobes converge into 6 to 10 major collecting ducts that have openings at the nipple and connect to the outside. Each of these major ducts arborizes back from the nipple and forms a lobe or segment of glandular tissue that is supported by surrounding connective or stromal tissue. The distribution of lobes is not even as there is a preponderance of glandular tissue in the upper outer quadrant of the breast. Beneath the nipple openings, the lactiferous sinus is visible. The lactiferous sinus is a slight dilation of the ampullary portion of the major duct. The major ducts that converge below the nipple and drain each segment are 2mm in diameter. Each duct drains a lobe made up of 20 to 40 lobules. Each lobule contains 10 to 100 alveoli or acini. Each lobule also consists of branching ducts that divide into sub segmental structures and terminate in the terminal duct lobular unit. The terminal duct lobular unit consists of the terminal duct and the acinus. The glandular tissue and ducts are surrounded by fat and supported by Cooper's ligaments, which are connective tissue elements that arise from stromal tissue and attach to the prepectoral fascia and dermis and support and suspend the breast tissue⁽¹⁰⁾.

2.2 Breast pathology:

The vast majority of the lesions that occur in the breast are benign. Much concern is given to malignant lesions of the breast because breast cancer is the most common malignancy in women in Western countries; however, benign lesions of the breast are far more frequent than malignant ones ⁽¹¹⁾.

2.2.1 Tumors of the breast:

2.2.1.1 Benign Breast Diseases:

2.2.1.1.1 Inflammatory and Related Lesions:

2.2.1.1.1.1 Mastitis:

These changes are a result of infectious agents; others do not have a well-understood etiology and may represent local reaction to a systemic disease, or a localized antigen-antibody reaction ⁽¹²⁾.

2.2.1.1.1.2 Acute Mastitis:

Acute mastitis usually occurs during the first 3 months postpartum as a result of breast feeding .Also known as puerperal or lactation mastitis, this disorder is a cellulitis of the interlobular connective tissue within the mammary gland, which can result in abscess formation and septicemia. It is diagnosed based on clinical symptoms and signs indicating inflammation ⁽¹³⁾.

2.2.1.1.1.3 Granulomatous Mastitis:

Granulomatous reactions resulting from an infectious etiology, foreign material, or systemic autoimmune diseases such as sarcoidosis and Wegener's granulomatosis can involve the breast many different types of organisms can cause granulomatous mastitis ⁽¹⁴⁾.

2.2.1.1.1.4 Foreign Body Reactions:

Foreign materials such as silicone and paraffin, which are used for both breast augmentation and reconstruction after cancer surgery, may cause a foreign body-type granulomatous reaction in the breast. Silicone granulomas ("siliconomas")

usually occur after direct injection of silicone in to the breast tissue or after extra capsular rupture of an implant ⁽¹⁵⁾.

2.2.1.1.1.5 Recurring Subareolar Abscess:

Recurring subareolar abscess (Zuska's disease) is a rare bacterial infection of the breast that is characterized by a triad of draining cutaneous fistula from the subareolar tissue; a chronic thick, pasty discharge from the nipple; and a history of multiple, recurrent mammary abscesses ⁽¹⁶⁾. The disease is caused by squamous metaplasia of one or more lactiferous ducts in their passage through the nipple, probably induced by smoking ⁽¹⁷⁾.

2.2.1.1.1.6 Mammary Duct Ectasia:

Also called periductal mastitis It is a disease of primarily middle-aged to elderly parous women who usually present with nipple discharge, a palpable subareolar mass, noncyclical mastalgia, or nipple inversion or retraction .Smoking has been implicated as an etiologic factor in mammary duct ectasia ⁽¹⁸⁾.

2.2.1.1.1.7 Fat Necrosis:

Fat necrosis of the breast is a benign nonsuppurative inflammatory process of adipose tissue. It can occur secondary to accidental or surgical trauma, or it may be associated with carcinoma or any lesion that provokes suppurative or necrotic degeneration, such as mammary duct ectasia and, to a lesser extent, fibrocystic disease with large cyst formation ⁽¹⁹⁾. It is characterized by anuclear fat cells often surrounded by histiocytic giant cells and foamy phagocytic histiocytes ⁽²⁰⁾.

2.2.1.1.1.8 Fibrocystic changes (FCCs):

Constitute the most frequent benign disorder of the breast. Such changes generally affect premenopausal women between 20 and 50 years of age ⁽²¹⁾. Although many other names have been used to describe this entity over the years, (including fibrocystic disease, cystic mastopathy, chronic cystic disease, mazoplasia, Reclus's

disease), the term “fibrocystic changes” is now preferred, because this process is observed clinically in up to 50% and histologically in 90% of women ⁽²²⁾.

2.2. 1.1.1.9 Cysts:

Are fluid-filled, round or ovoid structures that are found in as many as one third of women between 35 and 50 years old. Although most are subclinical “microcysts,” in about 20% – 25% of cases, palpable (gross) cystic change, which generally presents as a simple cyst. Cysts cannot reliably be distinguished from solid masses by clinical breast examination or mammography; in these cases, ultrasonography and fine needle aspiration (FNA) cytology, which are highly accurate, are used ⁽²³⁾.

2.2.1.1.1.10 Adenosis:

Adenosis of the breast is a proliferative lesion that is characterized by an increased number or size of glandular components, mostly involving the lobular units. Various types of adenosis have been described, of which sclerosing adenosis and microglandular adenosis ⁽²⁴⁾.

2.2.1.1.1.11 Metaplasia:

Apocrine metaplasia is characterized by the presence of columnar cells with abundant granular, eosinophilic cytoplasm and luminal cytoplasmic projections or apical snouts. These cells line dilated ducts or can be seen in papillary proliferations. They are more frequently found in younger women. All normal and metaplastic apocrine cells can be stained with gross cystic disease fluid protein. Atypical apocrine metaplasia should be diagnosed only when the nuclei of the apocrine cells display significant cytologic atypia ⁽²⁵⁾.

2.2.1.1.1.12 Epithelial Hyperplasia:

Epithelial hyperplasia is the most common form of proliferative breast disease, divided to ductal and lobular type ⁽²⁶⁾.

2.2.1.1.1.12.1 Ductal lesions:

The most important cytologic features of mild, moderate, or florid epithelial hyperplasia are an admixture of cell types (epithelial cells, myoepithelial cells, and metaplastic apocrine cells) and variation in the appearances of epithelial cells and their nuclei ⁽²⁶⁾.

2.2.1.1.1.12.2 Lobular Lesions:

Lobular neoplasia is most prevalent in perimenopausal women. It is a multifocal lesion, and many patients have lesions involving multiple quadrants of the breast, and increase the risk for the subsequent development of invasive carcinoma ⁽²⁷⁾.

2.2.1.1.1.13 Columnar Cell Lesions:

Columnar cell lesions of the breast represent a spectrum of lesions that have been encountered with increasing frequency in needle core breast biopsies because these lesions are commonly associated with microcalcifications and detected by mammographic screening ⁽²⁸⁾.

2.2.1.1.1.14 Intraductal Papilloma:

Intraductal papilloma is a discrete benign tumor of the epithelium of mammary ducts. It can arise at any point in the ductal system and shows a predilection for the extreme ends of the ductal system: the lactiferous sinuses and the terminal ductules ⁽²⁹⁾.

2.2.1.2 Proliferative Stromal Lesions:

2.2.1.3 Neoplasms:

2.2.1.3.1 Fibroadenoma:

Is the most common lesion of the breast; it occurs in 25% of asymptomatic women ⁽³⁰⁾. It is usually a disease of early reproductive life; the peak incidence is between the ages of 15 and 35 years. Conventionally regarded as a benign tumor of the breast, fibroadenoma is also thought to represent a group of hyperplastic breast lobules called “aberrations of normal development and involution. The lesion is a

hormone-dependent neoplasm that lactates during pregnancy and involutes along with the rest of the breast in perimenopause ⁽³¹⁾.

2.2.1.3.2 Adenoma:

An adenoma is pure epithelial neoplasm of the breast. This lesion is divided into tubular, lactating, apocrine, ductal, and so-called pleomorphic (i.e., benign mixed tumor) adenoma ⁽³²⁾.

2.2.2 Malignant Tumor of the Breast:

2.2.2.1 Noninvasive:

2.2.2.1.1 Intraductal carcinoma:

Carcinoma limited to the ducts (Ductal Carcinoma in situ DCIS) is reported in different age groups with increasing frequency, mainly attributable to the benefits of screening mammography ⁽³³⁾.

2.2.2.1.2 Lobular carcinoma in situ:

It is generally a non-palpable lesion diagnosed by mammography which may be located adjacent to fibrocystic changes or occurs concomitantly with infiltrative lobular carcinoma ⁽³³⁾.

2.2.2.2 Invasive:

2.2.2.2.1 Invasive ductal carcinoma:

This is the most common type exhibiting marked increase in dense fibrous stroma or desmoplastic response giving the tumour a hard consistency (Scirrhus). This type of cancer is usually associated with DCIS. The tumor margins are usually irregular ⁽³³⁾.

2.2.2.2.2 Invasive lobular carcinoma:

Arises from the terminal ductules of the breast lobule. This type tends to be bilateral and multi centric ⁽³³⁾.

2.2.2.2. 3 Mucinosa carcinoma:

Tends to occur in older patients and often produces large masses which give the tumor its soft consistency on palpation ⁽³³⁾.

2.2.2.2. 4 Medullary carcinoma:

This defined by (WHO) as a well-circumscribed carcinoma composed of poorly differentiated cells with scanty stroma and prominent lymphoid infiltration ⁽³³⁾.

2.2.2.2. 5 Papillary carcinoma:

A rare carcinoma in which invasive pattern is predominantly in the form of papillary structures. It may be adjacent to the nipple causing bloody or serosanguinous discharge ⁽³³⁾.

2.2.2.2. 6 Tubular carcinoma:

These tumors occur as small, firm, discrete masses. The (WHO) describes it as well differentiated carcinoma whose cells are arranged in regular well defined tubules typically lined by one epithelial layer and accompanied by abundant fibrostroma. Uncommon tumors, having characteristic cribriform pattern and are of the type seen more typically in the salivary gland ⁽³³⁾.

2.2.2.2. 2.7 Paget's disease of the nipple:

It is a specialized form of ductal carcinoma arising in the main secretory ducts and extend to involve the skin of the nipple and areola, which exhibit eczematous changes ⁽³³⁾.

2.2.3 Mixed Connective Tissue And Epithelial Tumors:

Fibro adenoma, Phyllodes Tumor (cystosarcoma phyllodes), Carcinosarcoma ⁽³⁴⁾.

2.3 Epidemiology of breast cancer:

Breast cancer is the most common malignancy in women ⁽³⁴⁾. Breast cancer is the most common cancer in women worldwide, with nearly 1.7 million new cases diagnosed in 2012 (second most common cancer overall). This represents about

12% of all new cancer cases and 25% of all cancers in women. It is the fifth most common cause of death from cancer in women ⁽²⁾.

In Sudan Data revealed that 74% of the women were < 50 years old or premenopausal. Invasive ductal carcinoma was the most common pathology (82%) and women presenting with stage III or higher tumors that had already metastasized, while ductal carcinoma in situ was the least prevalent (0.5%) finding ⁽³⁾.

2.4 Risk factors of breast cancer:

2.4.1 Sex:

Women are 100 times more likely to develop breast cancer than men ⁽³⁵⁾.

2.4.2 Age:

Increasing age is one of the strongest risk factors for breast cancer. Age is considered to be a likely surrogate for DNA damage accumulated during life ⁽³⁶⁾.

2.4.3 Affluence and lifestyle:

Breast cancer occurs more frequently in affluent and western populations ⁽³⁷⁾.

2.4.4 Family history:

Family history is an important and well-established breast cancer risk factor. Women with a mother, sister or daughter with breast cancer are, on average, at twice the risk of those with no affected first-degree relative ⁽³⁸⁾.

2.4.5 Endogenous oestrogens:

Postmenopausal women with high levels of circulating oestrogens (women with levels in the top 20%) have a two-fold increased risk of breast cancer compared with women with low levels of circulating oestrogens (women with levels in the bottom 20%) ⁽³⁹⁾.

2.4.6 Hormonal factors:

Factors such as reproductive history, menstrual history, menopausal status and exogenous hormone use are associated with breast cancer risk ⁽⁴⁰⁾.

2.5 Method of diagnosis of breast cancer:

2.5.1 Imaging Tests to Find Breast Cancer:

Different tests can be used to look for and diagnose breast cancer. If your doctor finds an area of concern on a screening test (a mammogram), or if you have symptoms that could mean breast cancer, you will need more tests to know for sure if it's cancer. Mammograms, Breast Ultrasound, Breast MRI Scans, Newer and Experimental Breast Imaging Tests ⁽⁴¹⁾.

2.5.2 Biopsy:

A biopsy is done when mammograms, other imaging tests, or a physical exam shows a breast change that may be cancer. A biopsy is the only way to know for sure if it's cancer ⁽⁴²⁾.

2.5.3 Fine needle aspiration (FNA) biopsy:

In an FNA biopsy, a very thin, hollow needle attached to a syringe is used to withdraw (aspirate) a small amount of tissue from a suspicious area. The needle used for an FNA biopsy is thinner than the one used for blood tests ⁽⁴³⁾.

2.5.4 Core needle biopsy:

A core biopsy uses a larger needle to sample breast changes felt by the doctor or seen on an ultrasound, mammogram, or MRI ⁽⁴⁴⁾.

2.5.5 Surgical (open) biopsy:

In rare cases, surgery is needed to remove all or part of the lump for testing. This is called a surgical or open biopsy. Most often, the surgeon removes the entire mass or abnormal area as well as a surrounding margin of normal breast tissue ⁽⁴⁵⁾.

2.5.6 Lymph node biopsy:

The doctor may also need to biopsy the lymph nodes under the arm to check them for cancer spread. This might be done at the same time as biopsy of the breast tumor ⁽⁴⁶⁾.

2.5.7 Biomarker-based methods:

Such as radioimmunoassay, immunohistochemistry, enzyme-linked immunosorbent assay (ELISA) and fluoroimmunoassay also cater to the diagnostic requirements for breast cancer ⁽⁴⁷⁾.

2.6 Breast Cancer treatment:

2.6.1 Chemotherapy:

It is known that chemotherapy can be helpful for many breast cancer patients ⁽⁴⁸⁾.

2.6.2 Oncoplastic surgery:

Breast-conserving surgery (lumpectomy or partial mastectomy) can often be used for early-stage breast cancers ⁽⁴⁹⁾.

2.6.3 Triple-negative breast cancer:

Since triple-negative breast cancers cannot be treated with hormone therapy or targeted therapy such as HER2 drugs, the treatment options are limited to chemotherapy ⁽⁵⁰⁾.

2.6.4 Targeted therapy drugs:

Targeted therapies are a group of drugs that specifically target gene changes in cells that help the cells grow or spread. New targeted therapies are being studied for use against breast cancer, including PARP inhibitors ⁽⁵¹⁾.

2.6.5 Supportive care:

There are trials looking at different medicines to try and improve memory and brain symptoms after chemotherapy ⁽⁵²⁾.

2.7 Survivin:

Survivin is a protein that inhibits apoptosis and regulates cell division ⁽⁵³⁾. Survivin plays an important role in the suppression of apoptosis by either directly or indirectly inhibits the activity of caspases. Survivin expression was not found in most normal adult human tissues when examined for reports investigating the role of survivin in cancer, human survivin expression has been reported in some normal

adult human tissues, including colonic mucosa ⁽⁵⁴⁾, placenta ⁽⁵⁵⁾, bone marrow and keratinocytes of the basal layer of the skin ⁽⁵⁶⁾. Over expression of survivin has been reported in almost all human malignancies including bladder cancer, lung cancer, breast cancer, stomach, esophagus, liver, ovarian cancers and hematological cancers ⁽⁵⁷⁾.

2.8 Survivin and breast cancer:

Boidot *et al.* (2000) showed that survivin expression might induce breast tumour proliferation by promoting genetic instability, the expression in breast cancer 70.7%-90.2% ⁽⁵⁸⁾.

Ambrosi *et al.*(1999) Survivin is expressed in most human cancers including that of colon, breast, lung, pancreas, prostate, stomach, esophagus, and in highgrade non-Hodgkin's lymphoma ⁽⁵⁹⁾ .

Yamashita *et al.* (2002) showed that there was a significant differences between the high and low expression groups ($p < 0.000$), Patients with low survivin expression showed better disease-free survival than patients with high survivin expression ⁽⁶⁰⁾.

Shin-chi Yamashita *et al.* (2007) reported that patients with low survivin expression showed significantly better disease-free survival than patients with high survivin expression in stage I and II breast cancer ($p < 0.0001$) ⁽⁶¹⁾.

Yong-Gang *et al.* (2010) assessed the possibility of survivin as a prognostic molecule. For example, in a study of 275 patients with breast cancer (70), survivin mRNA was highly expressed in tissues from younger patients (<50) and in high-grade cancer tissues. Survivin demonstrates a strong, independent association with poor prognosis ⁽⁹⁾.

Chapter Three

Materials and Method

3.1 Material and Methods

3.1 Materials:

Archived tissue block obtained from breast samples previously diagnosed as breast cancer were selected randomly for this study.

3.2 Methods:

3.2.1 Study design:

This is a descriptive, retrospective laboratory based study aimed to detect the expression of survivin in breast cancer among Sudanese women patients using immunohistochemistry.

3.2.2 Study sampling and sample size:

(40) Paraffin blocks previously diagnosed as breast cancer and positive control for survivin were selected from Elrahama medical center according to samples availability. Patient identification (age, grad and diagnosis) were obtained from patients records.

3.2.3 Study area:

This study was conducted at Elrahama medical center during the period from January to April 2018.

3.2.4 Immunohistochemistry Staining:

The immunohistochemical procedure was done as follows: one section (3 μ m) from formalin-fixed, paraffin-embedded tumors using tissue micro array technique, were cut and mounted onto Stalvanized slides (Thermo). Following deparaffinization in xylene, slides were rehydrated through a graded series of alcohol and were placed in distilled water. Samples were steamed for antigen retrieval for Survivin using high PH (9) by water bath at 95C for 40 min. After washing with PBS for 3 min Endogenous peroxides activity were blocked with 3% hydrogen peroxide and methanol for 10 min, and After washing with PBS for 3 min then Slides were incubated with (100 μ L) of (rabbit monoclonal antibody (Survivin) Quartett),

against Survivin antigen for 30 min at room temperature in a moisture chamber. After washing with PBS for 3 min, binding of antibodies will be detected by incubating for 20 min with dextrin labeled polymer (Dako). Finally, the sections washed in three changes of PBS, followed by adding 3, 3 diaminobenzidinetetrahydrochloride (DAB) as a chromogen to produce the characteristic brown stain for the visualization of the antibody/enzyme complex for up to 5 min. After washing with distilled water for 3 min. Slides were counterstained with haematoxylin (MAYER'S) for one min and washed in running tap water for several minutes 7-10 (bluing), then dehydrate, cleaned, and mounting DBX. Each slide was evaluated with investigator then the results were confirmed by consultant histopathologist.

3.2.5 Data analysis:

The obtained results and variables arranged in standard master sheet, then analyzed using statistical package for social (SPSS) program. Frequencies, means and chi square tests were calculated.

3.2.6 Result interpretation:

All quality control measures were adopted; positive and negative control sections were used during immunohistochemical staining. Detection of more than 5 cells with cytoplasmic and nuclear reaction per one field considered as positive result.

3.2.7 Ethical consideration:

The Samples were collected after permission according to the laboratory guidelines and regulation.

Chapter Four

Results

4. Result

The study included forty samples previously diagnosed as breast cancer. The age of patients ranged between 30 to 71 years with mean age of 50 years, and standard deviation was 10.7.

Most of patients were less than 50 years old representing 23/40 (57.5%) and the remaining 17/40 (42.5%) were more than 50 years, as indicated in table (4.1).

The histological grade of the study samples includes 11/40 (27.5%) of grade II, and 29/40 (72.5%) of grade III, grade I was not present, as indicated in table (4.2).

Survivin expression was positive in all samples of malignant tumor 40/40 (100%), the expression of survivin was moderately expressed in grade II 11/40 (27.5%) and highly expressed in grade III 29/40 (72.5%). This result showed a significant association between survivin antigen expression and histological grade (p-value = 0.000) as indicated in table (4.3).

Regarding the age of patients, the age group that were less than 50 years, Survivin was moderately expressed in 7/23 (30.4%) and highly expressed in 16/23 (69.6%), more than 50 years group, Survivin was moderately expressed in 4/17 (23.5%) and highly expressed in 13/17 (76.5%). This result showed no association between survivin antigen expression and age (p-value = 0.508) as indicated in table (4.4).

Table (4.1): Distribution of age groups among study patients:

Age	Frequency	Percent
≤ 50 years	23	57.5
≥ 50 years	17	42.5
Total	40	100

Table (4.2): Distribution of histological grade among study samples:

Grade	Frequency	Percent
Grade I	11	27.5
Grade II	29	72.5
Total	40	100

Table (4. 3): Correlation between the expression of survivin in breast carcinoma and histological grading:

Grade	Expression		Total	P.Value
	Moderate	High		
Grade II	11 (27.5%)	0 (0.0%)	11 (27.5%)	0.000
Grade III	0 (0.0%)	29 (72.5%)	29 (72.5%)	
Total	11 (27.5%)	29 (72.5%)	40 (100.0%)	

Table (4. 4): Correlation between expression of survivin and patients age:

Age	Expression		Total	P.Value
	Moderate	High		
≤ 50 years	7 (17.5%)	16 (40%)	23 (57.5%)	0.508
≥ 50 years	4 (10%)	13 (32.5)	17 (42.5%)	
Total	11 (27.5%)	29 (72.5%)	40 (100.0%)	

Chapter Five

Discussion

5. Discussion

The present study included 40 samples of breast carcinoma stained by immunohistochemistry for survivin antigen expression .The study revealed that patient age less than 50 years old are more affected with breast carcinoma 23(57.5%) .This result was compatible with Elgaili, et al. (2010), who reported that common involved age by breast carcinoma was the age group of 30-50 years .

The expression of survivin antigen was positive in all study samples of breast carcinoma 40/40 (100%), this agree with Ambrosi *et al.*(1999) is reported Survivin is expressed in most human cancers including that the breast.

Survivin antigen expression in study group was increase with grading of breast carcinoma (p-value = 0.000). This was agreed with Shin-chi Yamashita, et al. (2007) reported the survivin expression increased with breast tumor grading highest in grade III.

The present study revealed there was no significant association between survivin expression antigen and patient age (p-value 0.508).This result is disagreed with Alkhala, et al. (2013) who reported that tumor Survivin expression increases with age.

Chapter Six

Conclusion

Recommendations

6. Conclusion and Recommendations

6.1 Conclusion:

This study concluded that:

- The expression of survivin antigen was positive in all breast carcinoma.
- Survivin expression is associated with tumor grading, but no association with patient age.
- Most breast carcinoma patients in this study appear less than 50 years old.

6.2 Recommendation:

- Similar studies should be carried in larger sample size.
- Survivin recommended to be used as diagnostic and prognostic marker with other breast cancer marker panels.

Reference

- (1). Gershenson DM, Sun CC, Iyer RB, Malpica AL, Kavanagh JJ, et al. Hormonal therapy for recurrent low-grade serous carcinoma of the ovary or peritoneum. *Gynecol Oncol.*(2012), 125: 661-666.
- (2). World Cancer Research Fund International. Breast Cancer Statistics Available at <http://www.wcrf.org/int/cancer-factsfigures/dataspecificcancers/breast-cancer-statistics> Cancer Research UK, (2012).
- (3). International Journal of Women's Health. Breast cancer burden in central Sudan Available at https://www.researchgate.net/publication/47756204_Breast_cancer_burden_in_central_Sudan. (2010).
- (4). American cancer society. breast cancer basics. Available at <https://www.cancer.org/cancer/breast-cancer/risk-and-prevention.html> (2018).
- (5). National Cancer Institute Services Visit NCI's website <http://www.cancer.gov>, (2018).
- (6). National Cancer Institute. Breast Cancer visit NCI's website at <http://www.cancer.gov/cancertopics/types/breast>. (2018).
- (7). Altieri DC, Marchisio, PC . Survivin apoptosis: an interloper between cell death and cell proliferation in cancer. *Lab Invest.*(1999), 79:1327-133
- (8). Ambrosini G, Adida C, Altieri, DC. A novel anti-apoptosis gene, survivin, expressed in cancer and lymphoma. *Nat Med.*(1997) 3: 917-921
- (9). Yong-Gang, Lv#, Fang , Yu#, Qing Yao, Jiang-Hao Chen, Ling Wang J Thorac Dis .The role of survivin in diagnosis, prognosis and treatment of breast cancer .(2010), 2: 100-110.
- (10). Harris, JR, Lippman, ME, Morrow, M, Hellman, S. Diseases of the Breast. Philadelphia, PA: Lippincott-Raven Publishers (1996)
- (11). Caleffi M, Filho DD, Borghetti K, et al. Cryoablation of benign breast tumors: evolution of technique and technology.(2004).13:397–407.

- (12). Merih Guray and Aysegul A. Sahin ., *Oncologist*,(2006).11;435-449 , DOI: 10.1634/theoncologist.11-5-4Availableat <http://www.TheOncologist.com/cgi/content/full/11/5/435>
- (13).Foxman,B,D'Arcy,H,illespie,B,etal.Lactation mastitis: occurrence and medical management among946 breast feeding women in the United States. *Am J Epidemiol*.(2002), 55:103–114
- (14). Erhan ,Y, Veral ,A, Kara ,E, et al. A clinicopathologic study of a rare clinical entitymimickingbreastcarcinoma: idiopathic granulomatous mastitis *Breast*.(2000), 9:52–56
- (15). van Diest, PJ, Beekman, WH, Hage ,JJ. Pathology of silicone leakage from breast implants. *J Clin Pathol*. (1998),51:493–497.
- (16). Passaro, ME, Broughan ,TA, Sebek ,BA, et al.. Lactiferous fistula. *J Am Coll Surg*.(1994), 178:29–32.
- (17).Donegan, WL.Common benign conditions of the breast. In: Donegan WLSpratt JS, eds.*Cancer of the Breast, Fifth Edition*. St. Louis, MO: Saunders (2002),67–110
- (18). Furlong ,AJ, al-Nakib, L, Knox ,WF, et al.(1994). Periductal inflammation and cigarette smoke. *J Am Coll Surg*.(1994),179:417–420.
- (19). Rosai,J, ed.*Breast*.In:Rosai and Ackerman's *Surgical Pathology, Ninth Edition*. Philadelphia Chapter 20. Mosby.(2004),1763–1876.
- (20).Pullyblank, AM, Davies, JD, Basten, J, et al. Fat necrosis of the female breast--Hadfield re-visited. *Breast*.(2001) ,10:388–391
- (21). Cole ,P, Mark Elwood, J, Kaplan, SD. Incidence rates and risk factors of benign breast neoplasms. *Am J Epidemiol*(1978),108:112–120.
- (22).Santen,RJ, Mansel ,R. Benign breast disorders. *N Engl J Med* (2005),353:275–285.

- (23). O'Malley, FP, Bane, AL.(2004). The spectrum of apocrine lesions of the breast. *Adv Anat Pathol* .(2004),11:1–9
- (24).Lee, K, Chan, JKC, Gwi ,E. Tubular adenosis of the breast: a distinctive benign lesion mimicking invasive carcinoma. *Am J Surg Pathol* (1996).20:46–54.
- (25). Tavassoli ,FA, ed.Benign lesions. In: *Pathology of the Breast,Second Edition*. Stamford, CT: Appleton & Lange, Chapter(1999),5. 115–204.
- (26).Koerner,FC.Epithelialproliferationsofductaltype.*SeminDiagnPathol*.(2004),21 :10–1
- (27).Page ,DL, Schuyler, PA, Dupont ,WD, et al. Atypical lobular hyperplasia as a unilateral predictor of breast cancer risk: a retrospective cohort study. *Lancet* .(2003), 361:125–129.
- (28).Schnitt ,SJ, Vincent-Salomon, A.Columnar cell lesions of the breast. *Adv Anat Pathol*.(2003). 10:113–124
- (29).Oyama, T, Koerner ,FC. Noninvasive papillary proliferations. *Semin Diagn Pathol* .(2004), 21:32–41.
- (30). El-Wakeel, H, Umpleby ,HC. Systematic review of fibroadenoma as a risk factor for breast cancer. *Breast*. (2003),12:302–307
- (31). Hughes ,LE, Mansel ,RE, Webster, DJT. Aberrations of normal development and involution (ANDI): a new perspective on pathogenesis and nomenclature of benign breast disorders. *Lancet*. (1987),2:1316–1319.
- (32).Silverberg ,SG, DeLellis, RA, Frable ,WJ, eds. The breast. In: *Principles and Practice of Surgical Pathology and Cytopathology, Third Edition*. New York: Churchill-Livingstone, Inc.(1997), 575–673.
- (33).TheWorldHealthOrganizationHistologicalTypingofBreastTumors—Second EditionDownloadedfrom<https://academic.oup.com/ajcp/articleabstract/78/6/806/1812564> by guest on 18 July 2018

- (34).Dr. j. g. azzopard el. The World Health Organization Histological Typing of Breast Tumors—Second Edition, December. (1982), Vol. 78. No.6
- (35).NationalBreastand Ovarian Cancer Centre. Published with minor amendments. ISBN Online: (2009),978-1-74127-141-6
- (36). Perou, CM, Sorlie, T, Eisen ,MB, et al. Molecular portraits of human breast tumours. Nature.(2000),406:747-52.
- (37). Sorlie, T, Tibshirani, R, Parker ,J, et al. Repeated observation of breast tumor subtypesindependentgeneexpressiondatasets.ProcNatlAcadSciUSA(2003),100:8418-23.
- (38).ACIM(AustralianCancerIncidenceandMortality)Books.Canberra:Australian Institute of Health and Welfare (AIHW).(2007).
- (39). Thomas, DB. Breast cancer in men. Epidemiol Rev.(1997),15:220-31.
- (40). Globocan. International Agency for Research on Cancer (IARC).(2005).
- (41). Oeffinger, KC, Fontham, ET, Etzioni ,R,et al. Breast cancer screening for womenataveragerisk:guidelineupdateFromtheAmericanCancer Society.JAMA.(2015), 314(15):1599-1614.
- (42).Joe,BN,Esserman,LJ.BreastBiopsy.UpToDate.Accessedatwww.uptodate.com/contents/breast-biopsyonSeptembe5.(2017).
- (43).Radiological Society of North America, Inc. Stereotactic Breast Biopsy. Accessedatwww.radiologyinfo.org/en/info.cfm?pg=breastbixronSeptember 5,2017.
- (44).Sung JS, Comstock CE. Chapter 15: Image-Guided Biopsy of Nonpalpable BreastLesions. In: Harris JR, Lippman ME, Morrow M, Osborne CK, eds. Disease of the Breast. 5th ed. Philadelphia, Pa: Lippincott Williams & Wilkins; 2014.
- (45).American Cancer Society medical information is copyrighted material. For reprintrequests,pleasecontactpermissionrequest@cancer.org.Last Medical Review: September 1, 2017 Last Revised: October 9, 2017

- (46). Cheng, B. Y. Development of a chemiluminescent immunoassay for cancer antigen 15-3. *Labeled Immunoass. Clin. Med.* 2016, 23, 1348–1351.
- (47). Dizdar O, Arsian C, Altundag K. Advances in PARP inhibitors for the treatment of breast. *Expert Opin Pharmacother.* 2015;16(18):2751-2758
- (48). National Cancer Institute. <https://www.cancer.gov/aboutcancer/treatment/clinicaltrials/search>. Accessed August 9, 2017.
- (49). Vliek SB, Meershoek-Klein Kranenbarg E, van Rossum AGJ et al. The efficacy and safety of the addition of ibandronate to adjuvant hormonal therapy in postmenopausal women with hormone-receptor positive early breast cancer. First results of the TEAM IIB trial (BOOG 2006-04). Abstract 2016 San Antonio Breast Cancer Symposium
- (50). Robson ME, Im SA, Senkus E et al. OlympiAD: Phase III trial of olaparib monotherapy versus chemotherapy for patients (pts) with HER2-negative metastatic breast cancer (mBC) and a germline BRCA mutation (gBRCAm). Abstract 2017 American Society of Clinical Oncology Annual Meeting.
- (51). Vliek SB, Meershoek-Klein Kranenbarg E, van Rossum AGJ et al. The American Cancer Society Last Medical Review: July 1, 2017 Last Revised: February 1, 2018.
- (52). Altieri DC, Marchisio PC: Survivin apoptosis: an interloper between cell death and cell proliferation in cancer. *Lab Invest*, 1999; 79:1327-1333.
- (53). Gianani R, Jarboe E, Orlicky D et al: Expression of survivin in normal, hyperplastic, and neoplastic colonic mucosa. *Human Pathol*, 2001; 32: 119-125
- (54). Shiozaki A, Kataoka K, Fujimura M et al: Survivin inhibits apoptosis in cytotrophoblasts. *Placenta*, 2003; 24: 65-76

- (55).Grossman D, MaNiff JM, Li F, Altieri DC: Expression of the apoptosis inhibitor, survivin, in nonmelanoma skin cancer and gene targeting in a keratinocyte cell line. *Lab Invest*, 1998; 79: 1121-1126.
- (56). Praveen Kumar Jaiswal*,**, Apul Goel** & R.D. Mittal* Survivin: A molecular biomarker in cancer Mittal**Indian J Med Res* 141, April 2015, pp 389-39
- (57).EffusionsLilachKleinberg,MSc,1ViviAnnFlørenes,PhD,1JahnM.Nesland,MD, PhD,1,2 and Ben Davidson, MD, PhD1,2 *Am J Clin Pathol* Survivin, a Member of the Inhibitors of Apoptosis Family, Is Down-Regulated in Breast Carcinoma 2007;128:389-397 38 DOI:10.1309/E899BG1282M5D505
- (58).Tanaka K, Iwamoto S, Gon G, Nohara T, Iwamoto M, Tanigawa N. expression of survivin and its relationship to loss of apoptosis in breast carcinomas. *Clin Cancer Res* 2000;6 : 127-34.
- (59)Ambrosini G, Adida C, Altieri DC: A novel anti-apoptosis gene, survivin, expressed in cancer and lymphoma. *Nat Med*, 1997; 3: 917-921.
- (60).Rodel F, Hoffmann J, Grabenbauer GG, Papadopoulos T, Weiss C, Gunther K, Schick C, Sauer R and Rodel C: High survivin expression is associated with reduced apoptosis in rectal cancer and may predict disease-free survival after preoperative radiochemotherapy and surgical resection. *Strahlenther Onkol* 178: 426-435, 2002.
- (61).Barnes N, Haywood P, Flint P, Knox WF and Bundred NJ:Survivin expression in *in situ* and invasive breast cancer related to COX-2 expression and DCIS recurrence. *Br J Cancer* 94:253-258, 2006.

Appendices

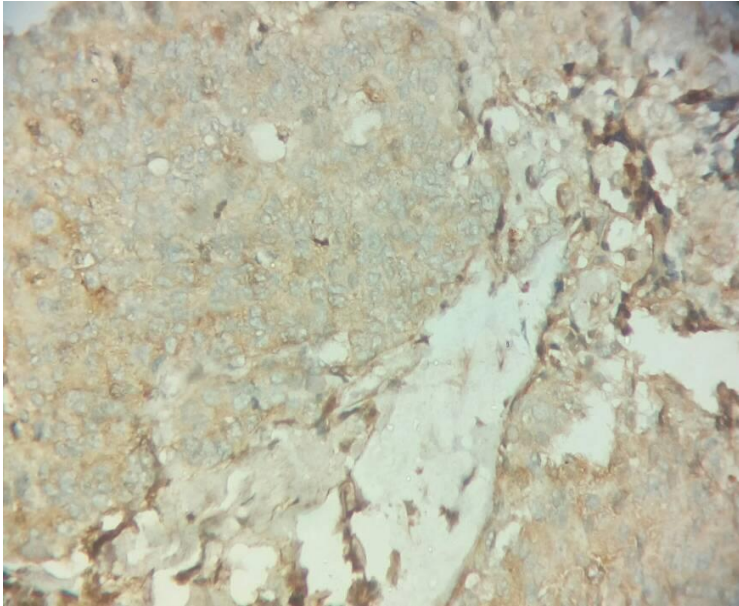
Appendix 1:

Materials and instrument used for processing and staining of the specimens include:

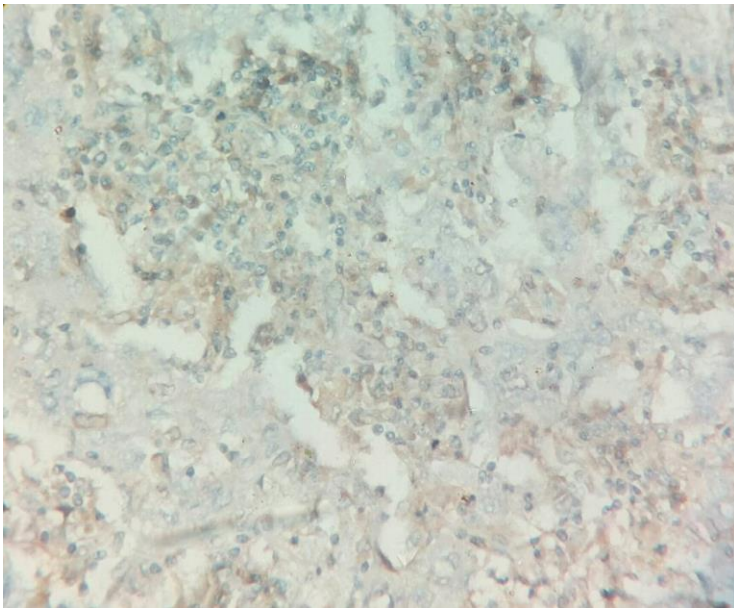
- Disposable gloves
- Microtome knife
- Positively charged slide
- Cover glass
- Dry oven
- Water bath
- Embedding center
- Coplin jar
- Humidity chamber
- Ethanol (100%, 90%, 70%, 50%)
- Mayer s haematoxylin (haematoxylin , DW,K or ammonium alum ,sodium iodated ,citric acid ,chororal hydrate)
- Reaction buffer
- Primary antibody (EBV)
- Tris EDTA buffer (PH9)
- Phosphate buffer saline (PH7.4)
- Peroxides blocker(3% hydrogen peroxide in methanol)
- Secondary anti body (dextran polymer conjugated secondary – HRP)
- DAB (3,3 di amino benzidin tetra hydrochloride)substrate solution
- Bluing Reagent (0.1MLi₂ CO₃ , 0.5 M Na₂CO₃)

- Xylene
- DPX mounting media

Microphotograph (4.1): high expression of Survivin in grade III



Microphotograph (4.2): moderate expression of Survivin in grade II



Cat. No.	2-SU044-10
Lot No.	351430
Quantity	3 ml

Antibody to Survivin

Host: Rabbit

Immunogen: Synthetic peptide corresponding to residues residues on the N terminus of human Survivin.

Subclass: IgG

Presentation: Prediluted antibody in TRIS, pH 7.4, containing < 0.1 % sodium azide.

Titre: Used in a corresponding system, vial is sufficient for 30 slides using 100 µl per slide. Incubate primary antibody for at least 60 min.

Assay system: IHC(p,f)

Fixation: 1) NOTOXhisto 2) Formalin

Treatments: Staining of formalin/paraffin tissues is enhanced by deparaffinization, rehydration and antigen retrieval. The recommended method is HIER (Heat Induced Epitope retrieval), including boiling of tissue sections in ProTaq Antigen Enhancer I, Cat# 401602092, for 20 min, followed by cooling at RT for 20 min. The optimal method of antigen retrieval must be determined in user's own system.

Reactivity: Human, others not tested.

Cell. Local.: Nuclear/Cytoplasmic

Control: Colon and Colon cancer

Storage: Aliquot and store at -20 °C. Do not use after expiration date indicated on the vial.

Errors excepted. This data sheet is a general information. The product attribute can diversify with changing Lot No as well as variations in tissue selection, tissue processing, antigen retrieval and detection systems. We do not take responsibility for any possible damage including personal injury, time or effort on economic loss caused by this product. Our warranty is limited to the price paid for the product. The product may only be used by authorized and skilled personnel. Non-application as prescribed in this data sheet leads to loss of all liability.

FOR IN VITRO USE, NOT FOR DIAGNOSTIC USE

Manufacturer:

BIOCYC GmbH & Co. KG, Am Mühlenberg 11, 14476 Potsdam; certified by ISO 13485:2012 and AC: 2012, ISO 9001:2008; Registry number: 60018916, 60018917

Distributed by:



Schichauweg 16, 12307 Berlin, Germany
 Tel.: ++49 (030) 765 925-0, Fax.: ++49 (030) 765 925-55
 E-Mail: info@quartett.com Internet: www.quartett.com